



USDA Food Safety and Inspection Service STEC Update

Philip Bronstein, PhD
USDA/FSIS/Science Staff
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Outline

- FSIS STEC Sampling
- Beef Veal Carcass Baseline Study
- Further Characterization of Strains
- Research Needs

Outline

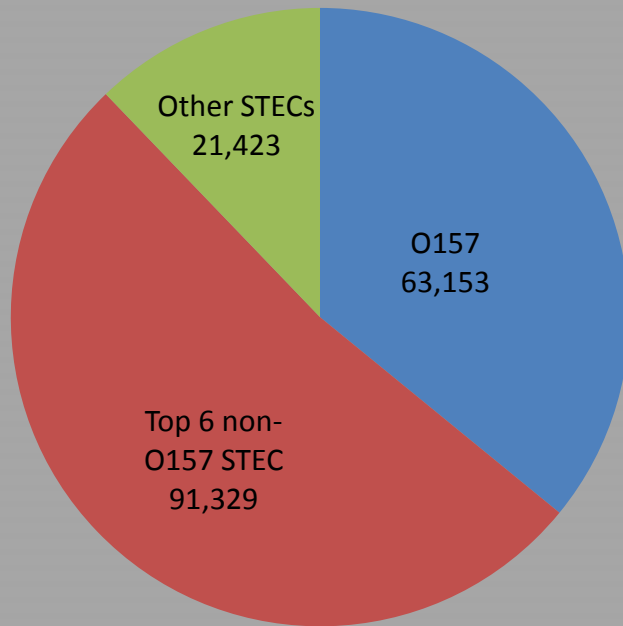
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Focus on STECs

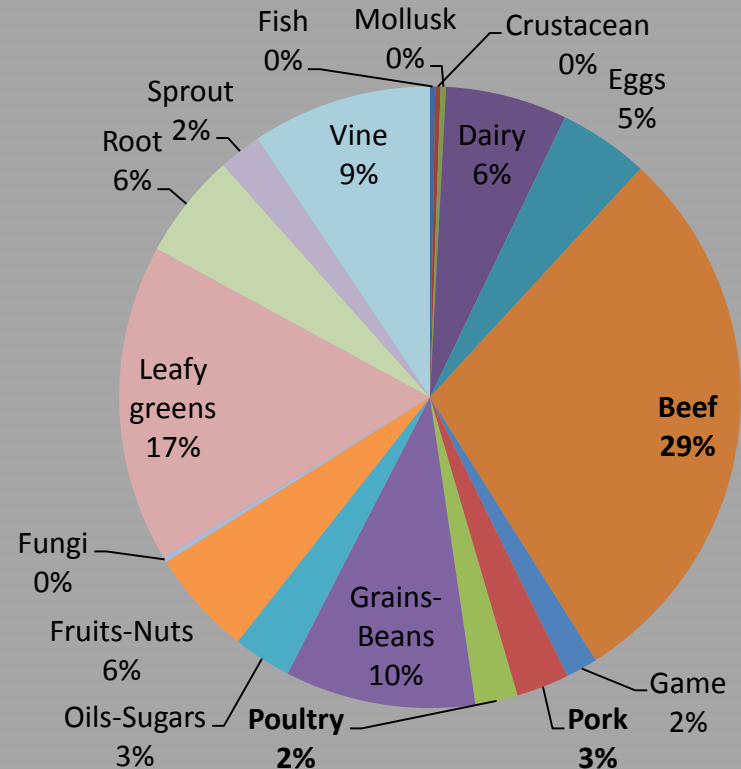
- In 1994 FSIS began testing raw beef products for *E. coli* O157:H7.
- Non-intact beef products containing *E. coli* O157:H7 were declared to be adulterated.
- In 2012 FSIS announced that the presence of additional shiga toxin-producing *E. coli* strains (STEC) in raw beef trim would render that product adulterated.
- On June 4, 2012 FSIS began sampling beef manufacturing trimmings for the 'top 6' non-O157 STECs
 - O-groups: O26, O45, O103, O111, O121, and O145
 - Strain must also include the virulence genes *eae* and *stx*

Focus on STECs

Foodborne related illnesses associated with STEC



STEC attribution to commodity





STEC Test Results

(June 4, 2012 through April 27, 2014)

Raw Ground Beef Components (RGBC)			
Serotype	Trim Verification	Follow-up to RGB Positive at Supplier	Follow-up to RGBC Positive
<i>E. coli</i> O157:H7	0.42% (22/5249)	0.81% (5/618)	0.54% (8/1476)
Total non-O157 STECs	0.70% (34/4887)	1.42% (7/492)	2.03% (26/1279)
Number of non-O157 STECs isolated from positive samples			
O26	9	1	11
O45	1	0	3
O103	18	2	21
O111	6	2	1
O121	0	0	0
O145	2	2	2



Beef/Veal Breakdown

Raw Beef Trim				
Source	Serotype	Trim Verification	Follow-up to RGB Positive at Supplier	Follow-up to RGBC Positive
Beef	<i>E. coli</i> O157:H7	0.37% (19/5162)	0.19% (1/537)	0.16% (2/1247)
	Total non-O157 STECs	0.58% (28/4808)	0.47% (2/425)	0.81% (9/1116)
	Number of non-O157 STECs isolated from positive samples			
	O26	9	1	4
	O45	0	0	0
	O103	16	1	5
	O111	4	0	0
	O121	0	0	0
	O145	1	0	0
Veal	<i>E. coli</i> O157:H7	3.57% (3/84)	4.94% (4/81)	2.64% (6/227)
	Total non-O157 STECs	7.89% (6/76)	7.46% (5/67)	10.43% (17/163)
	Number of non-O157 STECs isolated from positive samples			
	O26	0	0	7
	O45	1	0	3
	O103	2	1	16
	O111	2	2	1
	O121	0	0	0
	O145	1	2	2

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- FSIS STEC Sampling
- **Beef Veal Carcass Baseline Study**
- Further Characterization of Strains
- Research Needs



Purpose

- To gain information on the effectiveness of sanitary dressing post-hide removal and slaughter interventions pre-chill in beef/veal slaughter establishments;
- To provide process control criteria for beef/veal slaughter establishments;
- To estimate the prevalence and quantitative level of pathogenic organisms;
- To estimate the presence and quantitative levels of indicator organisms

Sample Collection Details

- Approximately, 4000 samples will be collected during both shakedown and the actual study from 133 beef and veal slaughter establishments.
- Swabs are being collected from the carcasses of all subclasses:
 - Beef (steers, heifers, cows, bulls, stag, and dairy cows)
 - Veal (heavy calves, bob veal, non-formula fed veal and formula fed veal)



Sampling Locations & Microbiological Targets

- Samples are being collected at two locations in the slaughter process:
 - “Post-hide removal”: (immediately after hide removal, before evisceration, and prior to the application of any interventions)
 - “Pre-chill”: at least 1 to 5 minutes after the last intervention has been applied and no more than one hour in the hotbox)
- When possible “Post-hide” and “Pre-chill” samples will be taken from leading and lagging sides of the same carcass
- Microbiological targets include:
 - Pathogens - *E. coli* O157:H7, non-O157 Shiga-toxin producing *E. coli* (STEC) and *Salmonella*
 - Indicator organisms - Generic *E. coli*, Total Aerobic Bacteria, Enterobacteriaceae and coliforms

Preliminary Shakedown Data

- Samples scheduled 798 (399 at Post-HR and 399 at Pre-Chill)
- Samples collected 664 (332 at Post-HR and 332 at Pre-Chill)
- Samples analyzed 620 (310 at Post-HR and 310 at Pre-Chill)

Distribution of Samples by Beef Subclass

Beef Subclasses	Samples by Subclass	Percent of Total Samples
Beef Carcasses		
Cow	38	12.3%
Steer	110	35.5%
Bull	9	2.9%
Dairy Cow	61	19.7%
Heifer	51	16.5%
Veal Carcasses		
Heavy Calf	9	2.9%
Non-formula fed Veal	1	0.3%
Formula-fed Veal	14	4.5%
Bob Veal	17	5.5%
Total	310	100%

Salmonella Data

Post-hide Removal

- *Salmonella* Percent Positive (60/309) 19.42%
- Serotype distribution

Serotype	Count	Percent
Montevideo	12	30.0%
Anatum	6	15.0%
Mbandaka	5	12.5%
Newport	4	10.0%
Agona	2	5.0%
Kentucky	2	5.0%
Muenchen	2	5.0%
Muenster	2	5.0%
Typhimurium	2	5.0%
Altona	1	2.5%
I 4,[5],12:i:-	1	2.5%
Infantis	1	2.5%
Total	40	100%

Pre-chill

- *Salmonella* Percent Positive (10/310) 3.22%
- Serotype distribution

Serotype	Count	Percent
Newport	2	22.2%
Typhimurium	2	22.2%
Agona	1	11.1%
Heidelberg	1	11.1%
Kentucky	1	11.1%
Montevideo	1	11.1%
Not Reportable	1	11.1%
Total	9	100%



Positive Results

Table 1. BVCBS Positive Sample Results

	Number of samples (includes both Post-hide and Pre-chill)	Number of Post-hide removal samples	Number of Pre-chill samples
Total Number of Analyzed Samples	620	310	310
Positive for Salmonella	60	51	9
Positive for E. coli O157:H7	2	2	0
Positive for non-O157 STEC	25	16	9
Positive for both Salmonella and E. coli O157:H7	0	0	0
Positive for both Salmonella and non-O157 STEC	9	8	1
Positive for both E. coli O157:H7 and non-O157 STEC	0	0	0
Positive for Salmonella, E. coli O157:H7, and non-O157 STEC	1	1	0

Table 2: Breakdown of BVCBS Positive STEC Serogroups

Month	Project	<i>E. coli</i> Serogroups							TOTAL
		O157:H7	O26	O45	O103	O111	O121	O145	
January	S52_PSTHR	2	1		2	1			6
	S52_PRECH					1			1
February	S52_PSTHR	1	4		4	2	1		12
	S52_PRECH		2		1	2			5
March	S52_PSTHR		7		7	1		1	16
	S52_PRECH		1		3				4
TOTALS	S52_PSTHR	3	12	0	13	4	1	1	34
	S52_PRECH	0	3	0	4	3	0	0	10
GRAND TOTALS		3	15	0	17	7	1	1	44



Beef/Veal Breakdown

Subclass	Post-Hide Removal Samples										
	<i>Salmonella</i>				<i>E. coli</i> O157:H7				non-O157 STECs		
	Number of Positives	Number Analyzed	Percent Positive		Number of Positives	Number Analyzed	Percent Positive		Number of Positives	Number Analyzed	Percent Positive
Beef Carcasses											
Beef Cow	10	38	26.3%		0	38	0.0%		3	38	7.9%
Bull	0	9	0.0%		0	9	0.0%		2	9	22.2%
Dairy Cow	24	61	39.3%		0	61	0.0%		3	61	4.9%
Heifer	7	51	13.7%		0	51	0.0%		3	51	5.9%
Steer	14	110	12.7%		2	110	1.8%		6	110	5.5%
Total Beef	55	269	20.4%		2	269	0.7%		17	269	6.3%
Veal Carcasses											
Bob Veal	4	17	23.5%		0	17	0.0%		4	17	23.5%
Formula-fed Veal	0	14	0.0%		0	14	0.0%		3	14	21.4%
Heavy Calf	1	9	11.1%		1	9	11.1%		0	9	0.0%
Non-formula fed Veal	0	1	0.0%		0	1	0.0%		1	1	100.0%
Total Veal	5	41	12.2%		1	41	2.4%		8	41	19.5%



Beef – Veal Breakdown

Subclass	Pre-Chill Samples										
	Salmonella				E. coli O157:H7				non-O157 STECs		
	Number of Positives	Number Analyzed	Percent Positive		Number of Positives	Number Analyzed	Percent Positive		Number of Positives	Number Analyzed	Percent Positive
Beef Carcasses											
Beef Cow	1	38	2.6%		0	38	0.0%		0	38	0.0%
Bull	0	9	0.0%		0	9	0.0%		0	9	0.0%
Dairy Cow	3	61	4.9%		0	61	0.0%		1	61	1.6%
Heifer	0	51	0.0%		0	51	0.0%		0	51	0.0%
Steer	2	110	1.8%		0	110	0.0%		4	110	3.6%
Total Beef	6	269	2.2%		0	269	0.0%		5	269	1.9%
Veal Carcasses											
Bob Veal	3	17	17.6%		0	17	0.0%		3	17	17.6%
Formula-fed Veal	1	14	7.1%		0	14	0.0%		2	14	14.3%
Heavy Calf	0	9	0.0%		0	9	0.0%		0	9	0.0%
Non-formula fed Veal	0	1	0.0%		0	1	0.0%		0	1	0.0%
Total Veal	4	41	9.8%		0	41	0.0%		5	41	12.2%

Timeline

- Shakedown is complete
 - Started 2nd Quarter FY2014 (January)
 - Ended April 3, 2014

- Actual Study start date
 - 4th Quarter FY2014 (July)
 - Will last for 12 months



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Research Objectives

- Identification of STECs in beef including serogroups that are not screened for during routine FSIS testing,
- Identification of co-occurrence of FSIS adulterant STECS and/or potentially pathogenic STECs in beef and
- Identification of virulence factors associated with FSIS adulterant STECS and/or potentially pathogenic STECs in beef

Procedures

- Enrichment broths of samples collected during routine FSIS sampling of beef products were blinded and sent to ARS US Meat Animal Center (USMARC) in Clay Center
 - Portions of the *E. coli* O157:H7 BAX[®] MP screen-positive raw beef (ground and trim) enrichment broths collected prior to the start of the non-O157 STEC testing program
 - Portions of the *stx* BAX[®] Real-time PCR Screening positive raw beef (ground and trim) enrichment broths collected after the start of the non-O157 STEC Testing programs
 - BAX negative raw beef samples enrichment broths were also sent to ARS.

Procedures

- ARS analyses include screening the enrichment broths for:
 - FSIS adulterant STECs,
 - Shiga toxin genes (*stx*),
 - intimin genes (*eae*)
- ARS also attempted to culture suspected STECs from enrichments that screened positive (for any of the above). Cultured isolates analyzed for:
 - O group
 - Shiga toxin genes (*stx*),
 - intimin genes (*eae*),
 - potential virulence factors

Results (To Date)

- ARS analyzed 369 broths submitted by FSIS.
 - 206 O157:H7 screen positive by FSIS
 - 163 O157:H7 screen negative by FSIS
- When ARS analyzed these broths, sixty eight of these broths (18%) screened positive for O157:H7 (BAX[®] MP).

Results (To Date)

- Two broths contained non-O157 adulterants STECs (one with O45 and a second with O103).
 - These adulterants possessed both the Shiga toxin gene (*stx+*) and intimin adherence gene (*eae+*).
 - The broth containing the O45 adulterant screened BAX[®] MP positive.
 - The broth containing the O103 adulterant screened BAX[®] MP negative.
 - No other adulterant STECs were culture confirmed from these broths.

Results (To Date)

- An additional non-adulterant STEC containing both the Shiga toxin gene (*stx+*) and intimin gene (*eae+*) was isolated from each of two other broths.
 - These isolates were identified as O5 and O74.
 - No adulterant *E. coli* were identified in either broth.
- Twenty broths contained non-O157 *E. coli* with Shiga toxin genes (*stx+*) but no intimin (*eae-*).
- Co-occurrence of O157:H7 and non-O157 adulterant STECs was not confirmed in any broths.
- Twenty broths contained non-O157 *E. coli* with Shiga toxin genes (*stx+*) but no intimin (*eae-*).

Preliminary Conclusions

- Potentially pathogenic STECs other than the current list of FSIS adulterant STECs are present in raw beef.
 - If these STECs are associated with severe human illness, then FSIS may consider expanding its list of adulterant STECs.
- The presence of an adulterant STEC can not be positively correlated to the presence of other pathogenic STECs.
 - The lack of consistent co-occurrence of STECs suggests that it may not be possible to identify a subset of STECs that can be used as indicator organisms for the entire suite of potentially pathogenic STECs.

Future

- Following the analysis of 1000 FSIS beef broths by ARS, the associated data will be analyzed. Possible conclusions include:
 - Additional research on this topic may be warranted
 - Publication in peer-reviewed journal
 - Changes to Agency approaches for the analysis of adulterant STECs in beef may be warranted.
 - Refinement of FSIS adulterant STEC definition

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FSIS Research Priorities

The screenshot shows a web browser window displaying the FSIS Research Priorities page. The browser title is "Food Safety Research Priorities - Windows Internet Explorer provided by FSIS". The address bar shows the URL: <http://www.fsis.usda.gov/wps/portal/fsis/topics/science/food-safety-research-priorities>. The page header includes the USDA logo, "Food Safety and Inspection Service", and "United States Department of Agriculture". Navigation links include "About FSIS", "District Offices", "Careers", "Contact Us", "Ask Karen", "askFSIS", and "En Español". A search bar is labeled "Search FSIS". A main navigation menu has "Topics", "Programs & Services", "Newsroom", and "Forms". The current page is "Topics / Science / Food Safety Research Priorities".

Food Safety Research Priorities

The Food Safety and Inspection Service (FSIS) is pleased to share a listing of the top food safety research areas of interest.

While FSIS is not a research funding organization, it recognizes the importance of keeping abreast of the latest scientific endeavors as well as its role in promoting research in areas important to the FSIS mission. This listing supports three of the FSIS 2011-2016 Strategic Plan goals:

1. strengthen collaboration among internal and external stakeholders to prevent foodborne illness,
2. effectively use science to understand foodborne illness and emerging trends, and
3. implement effective policies to respond to existing and emerging risks.

These priorities are presented as suggestions for researchers interested in pursuing food safety objectives that are relevant to FSIS regulated products. This list of research areas of interest may be useful to researchers who are preparing grants for submission to agencies that fund food safety research (e.g., USDA National Institute of Food and Agriculture (<http://www.nifa.usda.gov>), National Institutes of Health (<http://www.nih.gov>), <http://www.grants.gov>), or researchers with resources to conduct such research.

While FSIS is extremely interested in these research areas, this interest does not imply that the data and/or technologies generated by this research will be endorsed by FSIS.

This list represents FSIS' current assessment of priority research that will help further its public health mission; the list will be updated biannually. We encourage researchers to contact Dr. John Johnston by email (John.Johnston@fsis.usda.gov) or at (202) 365-7175 with questions. We also welcome information about research on related topics not currently listed here.

Select "For More Information" for any of the topics in the table below for a more detailed description and suggested research studies.

Research Priority Description
<ul style="list-style-type: none"> Investigate and/or develop emerging screening technologies to reduce time for detection. For More Information
<ul style="list-style-type: none"> Investigate and/or develop emerging screening technologies for enhanced subtype/virulence characterization of pathogens. For More Information
<ul style="list-style-type: none"> Investigate and/or develop emerging screening technologies to provide multi-analyte detection from a single analytical sample

Relevant FSIS Research Priorities

- Development of pathogen detection methodologies: Screening, enumeration, and characterization
 - Investigate and/or develop emerging screening technologies to reduce time for detection.
 - Investigate and/or develop emerging screening technologies for enhanced subtype/virulence characterization of pathogens.
 - Investigate and/or develop emerging screening technologies which are applicable to FSIS regulated products (meat, poultry, egg products and foods containing these products).
 - Develop or refine testing methods for *quantifying* target pathogens in meat, poultry and egg products.
 - Identify and evaluate alternative approaches to N60 sampling.
 - Identify and/or develop emerging technologies for real-time testing for higher levels of contamination prior to slaughter.
- Develop and evaluate interventions: Pre-harvest and at slaughter
 - Evaluate the potential effectiveness of pre-harvest interventions on finished products.
 - Determine the effectiveness of parallel and/or simultaneous application of more than one pre-harvest and/or post-harvest intervention as a control strategy.
 - Determine (validate) the effectiveness (log-reduction) of interventions used by industry to reduce levels of pathogens on FSIS regulated products.
 - Identify and/or develop pre- and post-harvest interventions to reduce levels of pathogens and chemical hazards for each class of veal (bob veal, non-formula-fed, formula-fed, and heavy calves).
- Conduct *ex post* evaluation of regulatory initiatives. Determine the presence and contributing factors for antimicrobial resistant strains in poultry and cattle.



Questions?